Correlation of two analytical methods for circulating tumor cells in peripheral blood of patients with primary breast cancer

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Background

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While the evidence for circulating tumor cells (CTCs) as a prognostic marker in metastatic breast cancer has been well established, there is still a lack of data in primary disease. In the SUCCESS A trial two different techniques for the detection of CTCs in early breast cancer were prospectively evaluated.

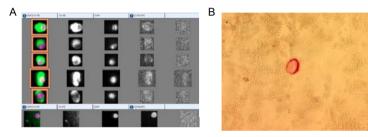


Figure 1:Sample pictures of CTCs, detected by CSS (A) and MICC (B). A: row 1 -5: CTCs; row 6: leukocyte.

Patients

Table1: Patients characteristics and CTC prevalence with the CellSearch System® (CSS) and the manual immuncytochemistry (MICC)

	CSS N	MICC N	CSS % pos	MICC
pT1	818	489	19,32	% pos 19,22
pT2-4	1159	751	22,69	22,24
pTx	17	9	17,65	33,33
pN0	680	445	19,41	20,22
pN1-3	1314	804	22,22	21,64
Gx	16	8	18,75	12,50
G1	98	59	14,29	23,73
G2	934	613	21,52	21,21
G3	946	569	21,78	20,91
HR -	569	371	21,79	22,10
HR +	1425	878	21,05	20,73
Her-2 x	50	29	18,00	20,69
Her-2 +	489	311	20,25	23,47
Her-2 -	1455	909	21,72	20,35
ductal	1600	1035	20,94	21,06
lobular	237	125	25,74	25,60
mixed	143	82	18,18	15,85
premen	830	529	20,00	21,55
postmen	1164	720	22,16	20,83

Materials & Methods

SUCCESS A compared FEC-Docetaxel vs. FEC-Docetaxel-Gemcitabine and 5 vs. 2 years of treatment with zoledronic acid in primary breast cancer patients and node positive or high-risk node negative disease.

Two different techniques to detect CTCs were prospectively evaluated in two consecutive, but comparable subgroups of the whole study population. In 3515 samples the CellSearch® System (CSS) (Veridex, Warren, USA) was used for CTC detection. Immunomagnetic enrichment with an EPCAMantibody was followed by labeling with monoclonal antibodies specific for cytokeratin (8, 18, 19) and leukocytes (CD45).

2165 samples were evaluated with a manual immunocytochemistry (MICC) protocol. Cytospins were prepared after mononuclear cell enrichment based on Oncoquick® centrifugation (greiner bio-one, Frickenhausen, Germany). Staining was performed with the monoclonal pancytokeratin antibody A45-B/B3 (Micromet, Munich, Germany) and the APAAP technique. Conventional light field microscopy (Axiophot; Zeiss, Oberkochen, Germany) was used for the detection of stained cells.

For both methods, the cut-off value for positivity was ≥ 1 CTC. All events were evaluated by two independent observers.

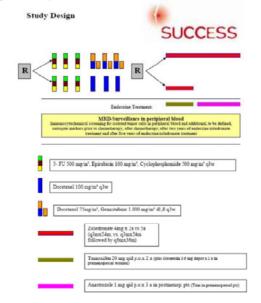
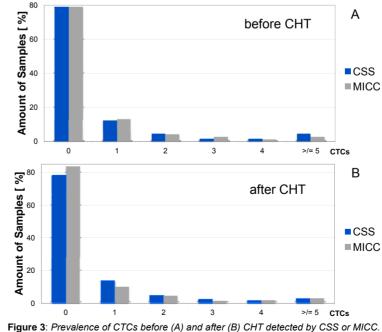


Figure 2: Study Design of the SUCCESS A trail.

Results

CTCs were examined in a total number of 3243 patients before and after chemotherapy (CHT). The two subgroups evaluated with one or the other method were well-balanced regarding clinical parameters as tumor size. grading, lymph node-status, hormone receptors and Her2. Furthermore there was no significant correlation between the CTC positivity and one of these clinical parameters using CellSearch® or the MICC, respectively (p > 0.05 using the chi square test each time). Before adjuvant CHT 21.3% (424 out of 1994) and 21.1% (264 out of 1249) of the patients were found positive for CTCs using CellSearch® or the MICC respectively, with a mean CTC level of 5.9 (range: 1 to 827) and 3.1 (range: 1 to 256). Immediately after CHT 21.9% (333 out of 1521) and 16.5% (151 out of 916) of the patients were positive for CTCs using CellSearch® or the MICC. The mean CTC level decreased to 3.0 (range: 1 to 124) and 2.1 (range: 1 to 23) in both analytical methods. Using CellSearch® there was a significant correlation between the presence of CTCs before CHT and disease progression (p = 0.0044), as well as survival (p = 0.0001), whereas the MICC did not predict any of these (p = 0.3143) and p = 0.0801 respectively; the chi-square test was used each time).



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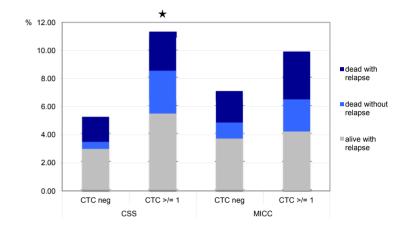


Figure 4: Percentage of patients who had a progress of their disease or died regarding the CTC-Positivity before CHT with one or the other method (median follow up 35 months). \star indicates a significant result ($p \le 0.05$ for disease progression and survival).

Conclusion

We found comparable prevalence of CTCs before and after adjuvant chemotherapy both with the CellSearch® System or the MICC. However, prognostic relevance could only be shown for CTCs detected with the CellSearch® System. This may be attributed to the high standardization and reproducibility of the automated system, as well as the additional CD45 counterstaining. According to our findings, the FDA approved CellSearch® System should be used as gold standard for CTC detection in future clinical trials.

Institutions

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